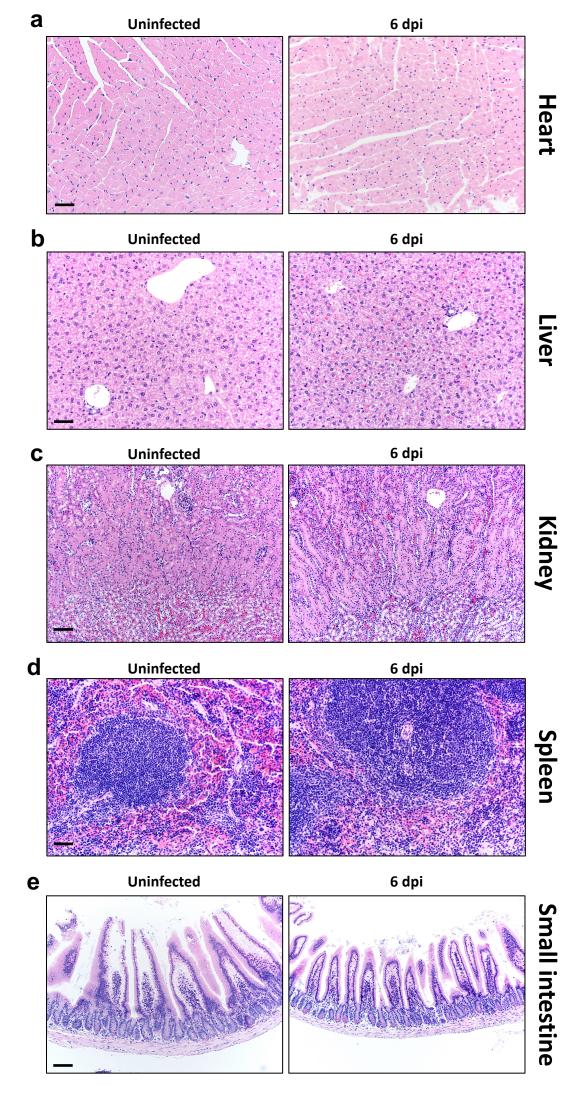


**Extended Data Figure 1** 

## Extended Data Figure 1. Infection of human respiratory cells and distribution of $SARS2-N501Y_{MA30}$ antigen in sinonasal cavity and brain.

**a**, Quantification of genomic RNA (gRNA), subgenomic RNA (sgRNA) and virus titers in Calu-3 cells at the indicated times after infection with 0.01 MOI of the indicated viruses. Data in **a** are geometric mean  $\pm$  geometric SD. **b**, Sinonasal cavity from young BALB/c mice infected with 5000 PFU of SARS2-N501Y<sub>MA30</sub>, H&E stain (top panels). Regions of respiratory epithelium and olfactory epithelium exhibited uncommon regional scattered (bottom-left panel) to localized SARS-CoV-2 nucleocapsid immunostaining, (bottom panels). Scale bar 90  $\mu$ m. OE: olfactory epithelium. **c**, Summary scores of nucleocapsid staining, as described in Methods (n=5). Data in **c** are mean  $\pm$  s.e.m. **d**, Brains from uninfected or infected young BALB/c mice at 6 dpi lacked overt lesions, H&E stain. Scale bar 45  $\mu$ m. **e**, Brains from uninfected mice or young BALB/c mice infected with 5000 PFU at 2,4, and 6 dpi revealed no SARS-CoV-2 nucleocapsid immunostaining. Scale bar 460  $\mu$ m.



**Extended Data Figure 2** 

## Extended Data Figure 2. Histological analysis of extrapulmonary tissue

Mice infected with 5000 PFU of SARS2-N501Y<sub>MA30</sub> were sacrificed at 6 dpi with 5000 PFU of SARS2-N501Y<sub>MA30</sub> and tissues were prepared for histological examination. Tissues from uninfected mice were analyzed in parallel.  $\mathbf{a}-\mathbf{e}$ , heart ( $\mathbf{a}$ ), liver ( $\mathbf{b}$ ), kidney ( $\mathbf{c}$ ), spleen ( $\mathbf{d}$ ), and small intestine ( $\mathbf{e}$ ) were studied. No overt, group-specific lesions were observed. Scale bars, 45 µm ( $\mathbf{a}$ ,  $\mathbf{b}$ ,  $\mathbf{d}$ ) and 90 µm ( $\mathbf{c}$ ,  $\mathbf{e}$ ), H&E stain. Two sections of each organ from 5 mice per group were evaluated.

**Extended Data Figure 3** 

## Extended Data Figure 3. Inflammatory mediators and immune effector cells in infected lungs.

Mice were infected with 1000 PFU SARS2-N501Y<sub>MA30</sub> to ensure survival until 6 dpi. **a,** Cytokine and chemokine transcripts were measured by qRT–PCR after isolation of RNA from the lungs of uninfected (0 dpi) and infected young BALB/c mice. Each lung was collected from an individual mouse. Mock (0 dpi), n=4; 2, 4, and 6 dpi, n=5. **b,** Quantification of immune cells (as gated in **e)** in the lungs (n = 3 for uninfected group; n = 4 and 5 for 4 and 6 dpi respectively) each lung was collected from an individual mouse). DC: dendritic cells; IMM: inflammatory monocytes/macrophages; PMN: polymorphonuclear cells. **c,** Representative FACS plots of IFN $\gamma$ +/TNF CD4 and CD8 T cells (as gated in **e)** after stimulation with indicated peptide pools in the lungs of young BALB/c mice at 6 dpi. **d,** Summary data for IFN $\gamma$  and TNF expression are shown (n=5). **e,** Gating strategy for identification of immune cells in lungs is shown. Data in **a, b, d** are mean  $\pm$  s.e.m

**Extended Data Figure 4** 

Extended Data Figure 4. Lymphopenia in SARS2-N501Y<sub>MA30</sub>-infected middleaged mice. Numbers of immune cells in the blood of young and middle-aged C57BL/6 mice at 3 and 6 days after infection with 5000 PFU of SARS2-N501Y<sub>MA30</sub> (n=4 for uninfected controls and middle-aged mice at 3 dpi; n=5 for middle-aged mice at 6 dpi and young mice at 3 and 6 dpi). PMN: Polymorphonuclear cells. P values determined by two-tailed Student's t test. P values indicated are determined from the corresponding time point compared to uninfected control of the same group. \*P=0.0003 (3 dpi vs uninfected, young C57BL/6; CD45\* cells); \*P=0.0194 (3 dpi vs uninfected, middle-aged C57BL/6, CD45+ cells); \*P=0.0102 (6 dpi vs uninfected, middle-aged C57BL/6, CD45+ cells); \*P=0.0046 (3 dpi vs uninfected, young C57BL/6; PMN); \*P=0.0304 (3 dpi vs uninfected, middle-aged C57BL/6, PMN); \*P=0.0318 (6 dpi vs uninfected, middle-aged C57BL/6, PMN); \*P<0.0001 (3 dpi vs uninfected, young C57BL/6; B cells); \*P=0.0264 (3 dpi vs uninfected, middle-aged C57BL/6, B cells); \*P=0.0049 (6 dpi vs uninfected, middle-aged C57BL/6, B cells); \*P=0.0025 (3 dpi vs uninfected, young C57BL/6; Total T cells); \*P=0.0262 (3 dpi vs uninfected, middle-aged C57BL/6, Total T cells); \*P=0.0051 (6 dpi vs uninfected, middle-aged C57BL/6, Total T cells); \*P=0.0115 (3 dpi vs uninfected, young C57BL/6; CD4+ T cells); \*P=0.0626 (3 dpi vs uninfected, middle-aged C57BL/6, CD4+ T cells); \*P=0.0061 (6 dpi vs uninfected, middle-aged C57BL/6, CD4+T cells); \*P=0.0001 (3 dpi vs uninfected, young C57BL/6; CD8+ T cells); \*P=0.0464 (3 dpi vs uninfected, middle-aged C57BL/6, CD8+ T cells); \*P=0.0054 (6 dpi vs uninfected, middle-aged C57BL/6, CD8+T cells). \* $P \le 0.05$ , \*\* $P \le 0.005$ , \*\*\* $P \le 0.0005$ , \*\*\*\* $P \le 0.0005$ , \*\*\*\* $P \le 0.0001$ . Data in **a** are mean ± s.e.m.